

Designing Molecular Swimmers

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The 21st Century Center-of-Excellence Program Exploring New Science by Bridging Particle-Matter Hierarchy The 4th COE Symposium, 28-30 June 2006

Swimming and Length Scale

• Simplified Dynamics

$$m\frac{d^2x}{dt^2} + V\frac{dx}{dt} = F(t)$$

F(t) is Periodic: interaction with medium

$$m \sim \mathbf{r}L^3 \qquad \mathbf{V} \sim \mathbf{h}L$$

Swimming and Length Scale

• Large Scales Inertial $t \to -t \Rightarrow F \to F$

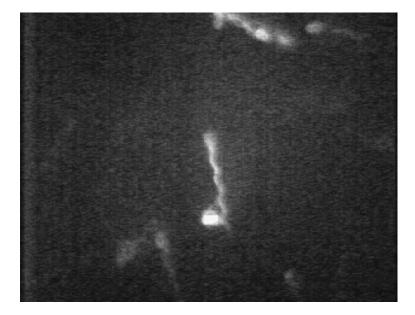
? constructive

• Small Scales Viscous $t \to -t \Rightarrow F \to -F$

? destructive

Bacterial Swimming

• *E. coli* driven by rotating flagella



L. Turner, W.S. Ryu, and H.C. Berg, *J. Bacteriol.* 182, 2793-2801 (2000)

• G.I. Taylor (1951) Swimming at low Reynolds number

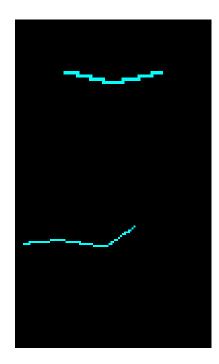
Swimming at Low Reynolds

• Purcell Swimmer (1976)

Using the smallest number of degrees of freedom:

- One, is not enough
 Scallop Theorem
- **Two**, will just do!

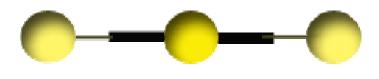
... but the passenger will get sea-sick!



Courtesy of Daniel B. Murray

Three-Sphere Swimmer

• Two Translational Degrees of Freedom



A. Najafi & R. Golestanian, PRE 69, 062901 (2004)

Swimming Velocity

• Simple Perturbative Analysis

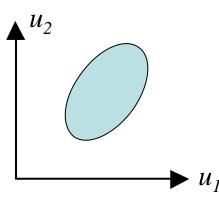
$$L_1 = \ell + u_1$$

$$L_2 = \ell + u_2$$

$$L_1 = L_1$$

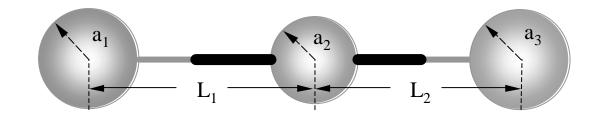
$$\overline{V} = \frac{7}{24} \frac{a}{\ell^2} \,\overline{(u_1 \dot{u}_2 - \dot{u}_1 u_2)}$$

• Geometric Interpretation



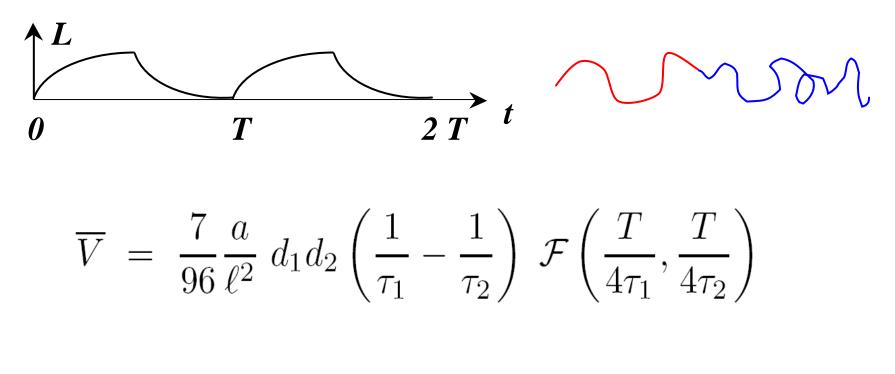
Harmonic Deformations

$$u_1(t) = d_1 \cos(\omega t + \varphi_1)$$
$$u_2(t) = d_2 \cos(\omega t + \varphi_2)$$



 $\overline{V} = \frac{7}{24} \frac{a}{\ell^2} d_1 d_2 \omega \sin(\varphi_1 - \varphi_2)$

Exponential Relaxation



$$\mathcal{F}(x,y) = \frac{1}{\sinh x \, \sinh y} \left[\frac{\sinh(x+y)}{(x+y)} - \frac{\sinh(x-y)}{(x-y)} \right]$$

Noisy Deformations

$$\overline{V} = \frac{7}{24} \frac{a}{\ell^2} \frac{1}{T} \int \frac{d\omega}{2\pi} i\omega \overline{[u_2(\omega)u_1(-\omega) - u_1(\omega)u_2(-\omega)]}$$

$$u_i(t) = \sum_n d_{in} \cos(\omega_n t + \varphi_{in})$$

$$\overline{V} = \frac{7}{24} \frac{a}{\ell^2} \sum_{n} \overline{d_{1n} d_{2n} \omega_n \sin(\varphi_{1n} - \varphi_{2n})}$$

Load-Velocity Response

• Force-Velocity Relation

$$\overline{V}(F) = \frac{7}{24} \frac{a}{\ell^2} \overline{(u_1 \dot{u}_2 - \dot{u}_1 u_2)} + \frac{F}{18\pi\eta a}$$

• Stall Force

$$F_s = -18\pi\eta a \overline{V}(0)$$

Mechanical Efficiency

• Power Consumption

$$\overline{\mathcal{P}} = 2\pi\eta a \left[\overline{\dot{u}_1^2} + \overline{\dot{u}_2^2} + \overline{(\dot{u}_1 + \dot{u}_2)^2} \right]^{18\pi n a \overline{V}^2}$$

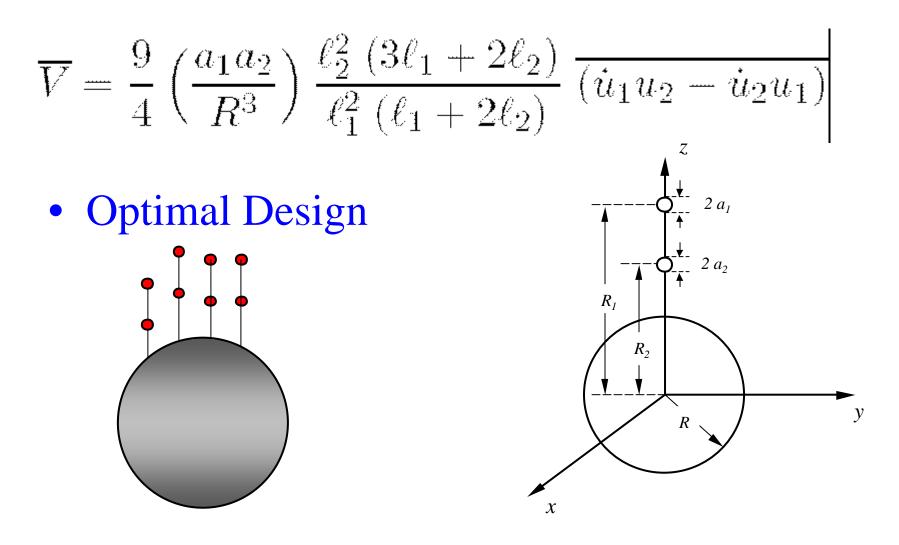
• Lighthill's Efficiency

$$\eta_{\rm L} = \frac{18\pi\eta a V^2}{\overline{\mathcal{P}}}$$

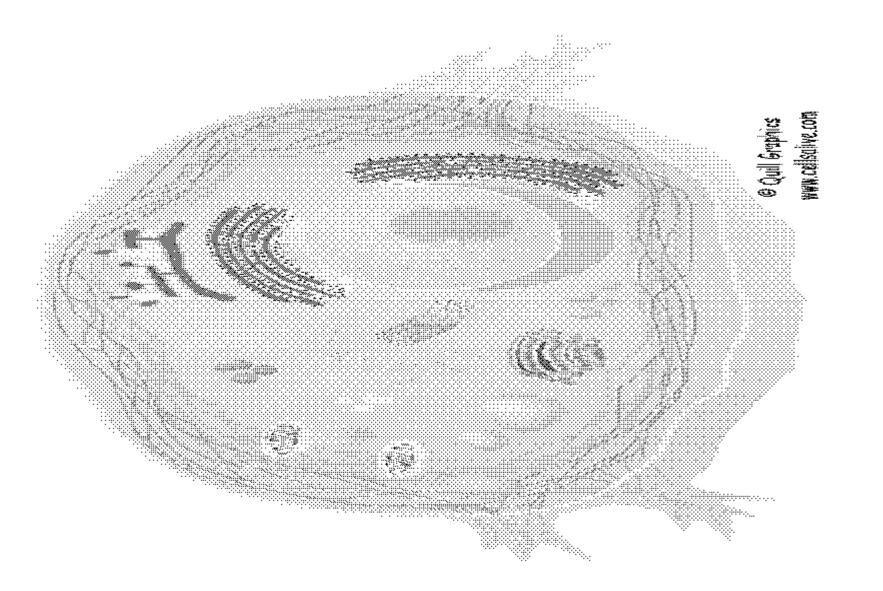
$$\eta_{\rm L} = \frac{49}{64} \frac{a^2}{\ell^4} \frac{\overline{(u_1 \dot{u}_2 - \dot{u}_1 u_2)}^2}{\overline{\dot{u}_1^2} + \overline{\dot{u}_2^2} + \overline{(\dot{u}_1 + \dot{u}_2)^2}}$$

Swimmer with Cargo

• Small deformations

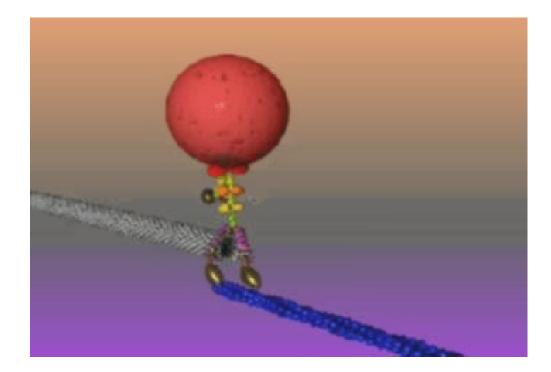


Transport System in the Cell



Molecular Motors

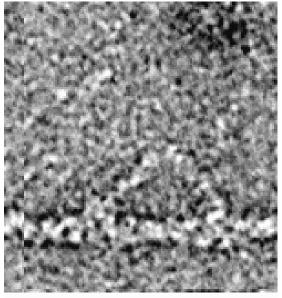
• Motion of myosin-V molecules on actin



Courtesy of Dr. George Langford

Molecular Motors

• Myosin-V: ``Electron Video Microscopy''



The Muscle Group, Leeds 2000

M L. Walker, S A. Burgess, J R. Sellers, F Wang, J.A. Hammer, J Trinick, & P J. Knight, Nature **405**, 804-807 (2000)

Molecular Motors

- Principal Characteristics
 - Nano-scale machines
 - Highly accurate despite Brownian agitation
 - Converting *chemical* energy directly to useful *mechanical* work

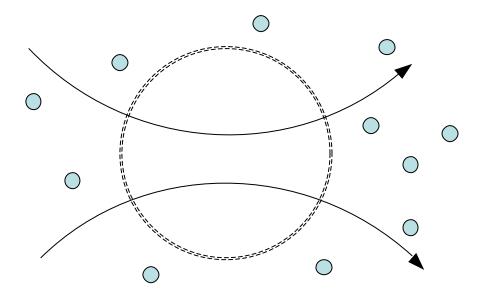
Can we design such a machine?

Phoretic Locomotion

• Osmio-phoresis

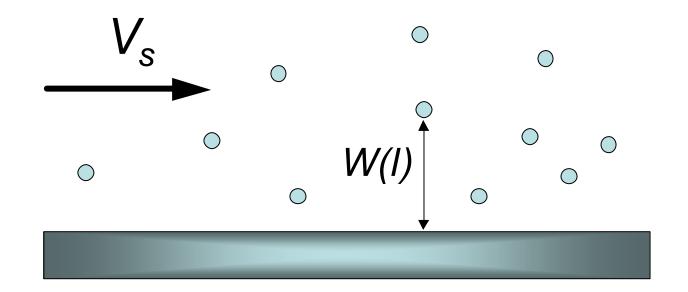
Motion of lipid vesicles under a gradient of sugar molecules

? Nardi, Sackmann & Bruinsma, PRL 82, 5168 (1999)

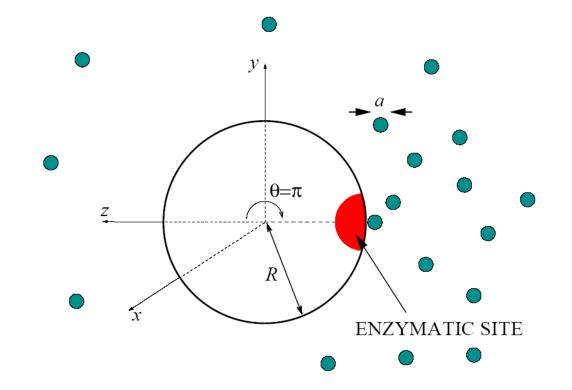


Phoretic Locomotion

- Diffusio-phoresis
- ? Derjaguin, Dukhin, Korotkova, Kolloidn. Zh. 23, 53 (1961)
- ? Anderson & Prieve, Sep. Purif. Methods 13, 67 (1984)



Reaction-Driven Propulsion



R. Golestanian, T.B. Liverpool & A. Ajdari, PRL 94, 220801 (2005)

Propulsion Velocity

$$v_0 = \frac{\ell}{\tau_{\rm f}} \qquad \ell = a \, \left(\frac{3 \, \lambda^2}{4 \, R^2}\right)$$

Diffusio-phoresis

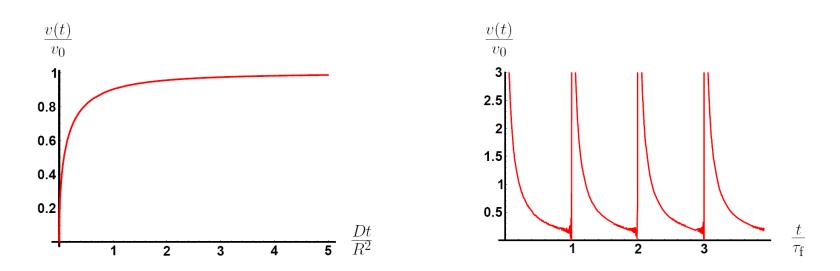
$$\lambda_D^2 = \int_0^\infty dl \ l \left[1 - e^{-W(l)/k_{\rm B}T} \right]$$

Osmio-phoresis

$$\lambda_O^2 = R^2 \left[\frac{\eta L_p / R}{2 + 20 \eta L_p / R} \right]$$

Time Response

Relaxation:Periodic particle release:



• Fluctuations: $\frac{(\Delta v)^2}{v_0^2} \sim D\tau_{\rm f}/R^2$

Reaction Kinetics

• Michaelis-Menten Rule

$$\mathcal{S} + \mathcal{E} \xrightarrow{k_1} \mathcal{S} \mathcal{E} \xrightarrow{k_2} \mathcal{P}_2 \mathcal{E} + \mathcal{P}_1 \xrightarrow{k_3} \mathcal{P}_1 + \mathcal{P}_2 + \mathcal{E}$$

$$\frac{1}{\tau_{\rm f}} = \left(\frac{k_2 k_3}{k_2 + k_3}\right) \frac{C_{\mathcal{S}}(\mathcal{E})}{K_M + C_{\mathcal{S}}(\mathcal{E})}$$

Michaelis constant: $K_M \equiv \frac{1}{k_1} \left(\frac{k_2 k_3}{k_2 + k_3} \right)$

Brownian Agitation

• Rotational Diffusion Time

$$\tau_R = 8\pi \eta R^3 / (k_{\rm B}T)$$

Directional persistence increases with size

Optimal Design

• Vesicle/bead of radius: $R = 2 \ \mu m$

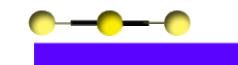
- Acetylcholinesterase: $1/\tau_{\rm f} \simeq 25000~{\rm s}^{-1}$
- Propulsion velocity: $v_0 \sim 0.5 \,\mu {\rm m/s}$
- Rotational diffusion time: 50 s

? Directional Locomotion

Conclusion

• Model A:

? externally controlled



- Model B:
 - ? reaction-driven

